Abietane Diterpenoids from the Cones of Larix kaempferi

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Three new abietane-type diterpenes, 7α , 15-dihydroxyabieta-8, 11, 13-trien-18-al (1); 15, 18-dihydroxyabieta-8, 11, 13-trien-18-al (1); 15, 18-dihydroxyabieta-8, 11, 13-trien-7-one (2); and 18-*nor*-4, 15-dihydroxyabieta-8, 11, 13-trien-7-one (3), were isolated from the cones of *Larix kaempferi*, together with three known diterpenes, abieta-8, 11, 13-trien-18-yl succinate, 16-*nor*-15-oxoabieta-8, 11, 13-trien-18-oic acid, and 7β -hydroxyabieta-8, 11, 13-trien-18-oic acid. The structures of **1**-3 were determined on the basis of chemical and spectral evidence.

Recently, we reported the isolation of 18-*nor*-abieta-8,-11,13-triene-4,15-diol and 18-*nor*-abieta-8,11,13-triene-4,7 α -diol from the cones of *Larix kaempferi* (Lamb.) Carr. (Pinaceae), together with two known diterpenes, abieta-8,11,13-triene-15,18-diol and abieta-8,11,13-triene-7 α ,18diol.¹

Further investigation of a CHCl₃ extract of the fresh cones of *L. kaempferi* furnished three new compounds (1–3), together with three known compounds. The known compounds were identified as abieta-8,11,13-triene-18-yl succinate,² 16-*nor*-15-oxoabieta-8,11,13-triene-18-oic acid,³ and 7β -hydroxyabieta-8,11,13-trien-18-oic acid⁴ by comparison of their physical, IR, ¹H and ¹³C NMR, and EIMS data with those already published. Compound 5 was previously isolated from the leaves of *L. kaempferi*.³ We now report the characterization of 1-3.



Compound **1** was assigned the molecular formula $C_{20}H_{28}O_3$, by HREIMS. Its IR spectrum indicated absorption bands for hydroxyl groups, an aldehyde group, and a benzene ring. The ¹H and ¹³C NMR spectra (Tables 1 and 2) showed signals for two tertiary methyl groups, two equivalent methyls of a hydroxylsopropyl group,¹ an aromatic ring characteristic for an abieta-8,11,13-triene, and an aldehyde group [δ_H 9.30 (1H, s); δ_C 206.2 (d)]. Acetylation of **1** afforded a monoacetate (**1a**). Except for the



Figure 1. HMBC (plain arrow) and key NOESY (dashed arrow) interactions of compound 1.

absence of a carboxyl group at C-18 and the presence of an aldehyde group, close resemblances were observed in the ¹H and ¹³C NMR spectra with analogous data of the known compound 7α,15-dihydroxyabieta-8,11,13-trien-18oic acid.⁵ The HMBC spectrum of 1 provided cross correlations shown in Figure 1, indicating that an aldehyde group should be placed at C-4. The configuration of the C-7 hydroxyl group of 1 was determined as pseudoaxial 7α based on the ¹³C NMR chemical shift values at C-5, C-6, C-7, and C-14 by comparison with those of 7β -hydroxyabieta-8,11,13-trien-18-oic acid.⁴ The unambiguous structure of 1 was determined from NOESY correlations between H-19 with H-20, and the aldehyde proton with the H-3 α and H-5 α protons (Figure 1). Therefore, compound 1 was determined to be 7α , 15-dihydroxyabieta-8, 11, 13trien-18-al.

Compound 2 was also established with the molecular formula C₂₀H₂₈O₃, by HREIMS. Its UV and IR spectra showed absorptions for hydroxyl groups, an α,β -unsaturated ketone, and a conjugated aromatic ring. The ¹H and ¹³C NMR spectra (Tables 1 and 2) showed signals for two tertiary methyl groups, a hydroxyisopropyl group, a primary hydroxyl group, a ketone group, and an aromatic ring characteristic of an abieta-8,11,13-triene. Acetylation of 2 afforded a monoacetate (2a) and a less polar product, diacetate (2b), in the ratio 2:1. The ¹H and ¹³C NMR spectral data of 2 resembled those of abieta-8,11,13-triene-15,18-diol,¹ except for the presence of a ketone group instead of a methylene group at C-7. This inference was supported by ¹H-¹H COSY, HMQC, HMBC, and NOESY experiments. The HMBC and NOESY data are shown in Figure 2. Therefore, compound 2 could be represented as 15,18-dihydroxyabieta-8,11,13-trien-7-one.

Compound **3** was assigned the molecular formula $C_{19}H_{26}O_3$, by HREIMS. The UV and IR spectra indicated absorptions for hydroxyl groups, an α,β -unsaturated ke-

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Table 1. ¹H NMR Spectral Data of Compounds 1, 1a, 2, 2a, 2b, and 3 in CDCl₃^a

proton	1	1a	2	2a	2b	3
Η-1α	1.49, m	1.50, m	1.53, m	1.57, ddd (13.3, 13.3, 4.0)	1.56, ddd (13.0, 13.0, 4.0)	1.57, m
H-1 β	2.35, ddd (13.0, 3.5, 3.5)	2.37. ddd (13.0, 3.5, 3.5)	2.33, ddd (13.0, 3.5, 3.5)	2.37, ddd (13.0, 3.5, 3.5)	2.34, ddd (13.0, 3.5, 3.5)	2.33, ddd (12.5, 2.5, 2.5)
Η-2α	1.84, m	1.84, m	1.76, m	1.78, m	1.77, m	1.85, d quintet (13.5, 3.0)
H-2 β	1.84, m	1.84, m	1.81, m	1.83, m	1.82, m	1.69, dddt (13.5, 13.5, 13.5, 3.0)
Η-3α	1.51, ddd (13.5, 13.5, 5.0)	1.48, m	1.59, ddd (13.3, 13.3, 3.8)	1.47, m	1.46, m	1.45, ddd (13.5, 13.5, 3.0)
H-3 β	1.38, dt (13.5, 3.0, 3.0)	1.40, ddd (13.5, 3.0, 3.0)	1.38, ddd (13.3, 3.8, 3.8)	1.47, m	1.46, m	1.93, ddd (13.5, 3.0, 3.0)
Η-5α	2.34, dd (13.0, 2.0)	2.34, dd (13.0, 2.0)	2.26, dd (12.7, 5.3)	2.21, dd (10.8, 7.0)	2.23, dd (10.8, 7.5)	2.13, dd (14.5, 4.0)
Η-6α	1.46, m	1.52, m	2.65, m	2.68, m	2.66, m	3.00, dd (18.0, 4.0)
H-6 β	2.06, m	2.08, ddd (14.0, 13.0, 4.3)	2.65, m	2.68, m	2.66, m	2.63, dd (18.0, 14.5)
$H-7\beta$	4.79, dd (4.5, 1.5)	5.97, dd (4.3, 1.5)				
H-11	7.26, d (8.5)	7.30, d (8.5)	7.34, d (8.5)	7.38, d (8.5)	7.35, d (8.5)	7.36, d (8.5)
H-12	7.38, dd (8.5, 2.0)	7.44, dd (8.5, 2.0)	7.68, dd (8.5, 2.0)	7.75, dd (8.5, 2.0)	7.52, dd (8.5, 2.0)	7.73, dd (8.5, 2.0)
H-14	7.46, d (2.0)	7.34, d (2.0)	7.99, d (2.0)	8.08, d (2.0)	7.98, d (2.0)	8.08, d (2.0)
H-15						
H-16	1.57, s	1.56, s	1.54, s	1.59, s	1.76, s	1.58, s
H-17	1.58, s	1.56, s	1.55, s	1.66, s	1.77, s	1.59, s
H-18	9.30, s	9.27, s	3.13, d (11.5) 3.46, d (11.5)	3.74, d (11.5) 3.84, d (11.5)	3.73, d (11.5) 3.83, d (11.5)	
H-19	1.17, s	1.17, s	0.93, s	1.03, s	1.02, s	1.30, s
H-20	1.19, s	1.21, s	1.25, s	1.28, s	1.26, s	1.21, s
C(15)OCOMe					2.05, s	
C(18)OCOMe		2.08, s		2.02, s	2.04, s	

^{*a*} Values were recorded at 500 MHz, δ in ppm, J (in parentheses) in Hz; assignments from ¹H-¹H COSY, HMQC, HMBC, and NOESY data.

Table 2.	¹³ C	NMR	Spectral	Data	of	Compounds	1,	1a ,	2,	2a,
2b, 3 (Cl	OCl ₃) ²	2								

carbon	1	1a	2	2a	2b	3
1	37.5 t	37.4 t	37.5 t	37.3 t	37.3 t	37.2 t
2	17.8 t	17.7 t	18.2 t	18.0 t	18.0 t	20.1 t
3	31.9 t	32.0 t	34.7 t	35.2 t	35.2 t	42.5 t
4	49.2 s	49.1 s	37.7 s	37.7 s	37.7 s	71.5 s
5	37.3 d	38.1 d	42.2 d	43.2 d	42.8 d	50.9 d
6	30.9 t	28.3 t	35.9 t	36.0 t	35.9 t	35.0 t
7	67.8 d	70.1 d	199.8 s	198.8 s	198.6 s	198.9 s
8	135.7 s	131.8 s	130.4 s	130.4 s	130.6 s	130.4 s
9	147.1 s	147.9 s	154.5 s	154.2 s	154.2 s	153.5 s
10	36.7 s	36.6 s	37.6 s	36.6 s	36.5 s	38.6 s
11	124.3 d	124.4 d	123.7 d	123.7 d	123.7 d	124.0 d
12	124.9 d	125.4 d	130.6 d	130.6 d	130.1 d	130.6 d
13	147.0 s	147.2 s	147.1 s	147.2 s	144.0 s	147.2 s
14	126.0 d	126.4 d	122.9 d	123.1 d	123.1 d	123.1 d
15	72.3 s	72.2 s	72.2 s	72.3 s	81.0 s	72.2 s
16	31.6 q	31.6 q	31.5 q	31.6 q	28.5 q	31.6 q
17	31.7 q	31.7 q	31.5 q	31.7 q	28.7 q	31.6 q
18	206.2 đ	205.6 đ	70.6 t	71.6 t	71.4 t	-
19	14.0 q	14.1 q	17.3 q	17.3 q	17.3 q	22.7 q
20	24.3 q	24.3 q	23.8 q	23.9 q	23.9 q	22.7 q
C(15)OCOMe			•		22.3 q	-
C(15)O <i>C</i> OMe					169.8 s	
C(18)OCOMe		21.5 q		20.9 q	21.0 q	
C(18)O <i>C</i> OMe		170.5 s		171.0 s	171.1 s	

 a Values were recorded at 125 MHz, δ in ppm; assignments from DEPT, HMQC, and HMBC experiments.

tone, and a conjugated aromatic ring. The ¹H and ¹³C NMR spectra (Tables 1 and 2) showed signals for two tertiary methyl groups, a hydroxyisopropyl group, a tertiary hydroxyl group, and an aromatic ring characteristic of an abieta-8,11,13-triene. Comparing the ¹³C NMR data of **3** with that of **2**, compound **3** shows a signal attributed to a quaternary oxygenated carbon [δ_C 71.5 (s)], while the C-18 signal of **2** [δ_C 70.6 (t)] was absent. Together with the molecular ion at m/z 302.1889 in EIMS, these data suggested that compound **3** was a new norabietatriene. The HMBC spectrum of **3** exhibited the cross correlations shown in Figure 3, indicating that two hydroxyl groups should be placed at C-4 and C-15, and a ketone group at C-7. In the NOESY spectrum (Figure 3), a significant



Figure 2. HMBC (plain arrow) and key NOESY (dashed arrow) interactions of compound 2.



Figure 3. HMBC (plain arrow) and key NOESY (dashed arrow) interactions of compound 3.

correlation was observed between the signals of H-20 and H-19 geminal to a hydroxyl group, indicative of a 1,3-diaxial relationship. Thus, compound **3** was characterized as 18-*nor*-4,15-dihydroxyabieta-8,11,13-trien-7-one.

Compounds 1-3 have not yet been reported in the literature. Although the ¹H and ¹³C NMR data of compound **2a** have been reported,⁶ some differences in the ¹³C NMR assignments were observed in the present study (Table 2).

Experimental Section

General Experimental Procedures. Melting points were determined on a Yanagimoto micromelting-point apparatus and are uncorrected. Optical rotations were measured using a JASCO DIP-1000 digital polarimeter. UV spectra were recorded on a Hitachi 150-20 spectrophotometer, and IR spectra were recorded using a Perkin-Elmer 1720X FTIR spectrophotometer. ¹H and ¹³C NMR spectra were obtained on Varian XL-300 and INOVA 500 spectrometers with standard pulse sequences, operating at 300 and 500 MHz, and 74.5 and 125 MHz, respectively. \widetilde{CDCl}_3 was used as solvent and TMS as internal standard. EIMS and HREIMS were recorded on a Hitachi 4000H double-focusing mass spectrometer (70 eV). Column chromatography was carried out over Si gel (70-230 mesh, Merck) and Cosmocil 75 C18-OPN (ODS, Nacarai Tesque), and MPLC was carried out with Si gel (230-400 mesh, Merck) and Cosmocil 40 C₁₈-PREP (ODS, Nacarai Tesque). Preparative HPLC was carried out using a TOSOHsystem equipped with a CCPM-prep pump, a SC-8020 system controller, and a TSK-GEL ODS-80Ts (21.5 × 300 mm) column. Fractions obtained from column chromatography were monitored by TLC (Si gel 60 HF₂₅₄). Preparative TLC was carried out on Merck Si gel PF_{254} plates (20 \times 20 cm, 0.5 mm thick).

Isolation of Compounds. Preliminary Si gel column chromatography of the CHCl₃ extract of the fresh cones of L. kaempferi has been reported previously, with separation into 10 fractions.¹ Rechromatography of fraction 8 (9.87 g), eluted with CHCl₃-EtOAc (2:1) from the preliminary Si gel column chromatography, over Si gel (200 g) with a solvent gradient from *n*-hexane-EtOAc (4:1) to 100% EtOAc afforded fractions a-k. Rechromatography of fraction f (169 mg), eluted from *n*-hexane–EtOAc (2:1), over ODS column with MeOH– H_2O (3:1) furnished 7β -hydroxyabieta-8,11,13-trien-18-oic acid, 39 mg, $[\alpha]^{23}_{D} + 21^{\circ}$ (*c* 0.41, EtOH).⁴ Fraction **g** (1.65 g), obtained from *n*-hexane–EtOAc (1:1), was repeatedly purified by ODS column chromatography with MeOH-H2O (4:1) to give successively compound 2 (120 mg) and compound 1 (8 mg). Rechromatography of fraction h (241 mg), obtained from *n*-hexane–EtOÅc (1:1), was purified using MPLC (ODS) with MeOH-H₂O (4:1), and HPLC with MeCN-H₂O (7:3) furnished compound 3 (20 mg).

Fraction **F** (2.17 g), collected from the early fractions in the rechromatography of fraction 9, was subjected to MPLC (Si gel). Elution with *n*-hexane–EtOAc (3:1) successively afforded two gummy residues from fractions 13–28 (**F-1**, 338 mg) and 29–48 (**F-2**, 195 mg), respectively. Rechromatography of **F-1** over an ODS column with MeOH–H₂O (4:1) furnished crude 16-*nor*-15-oxoabieta-8,11,13-trien-18-oic acid (12 mg), which was methylated by diazomethane etherate to afford a methyl ester (6 mg), identical in all respects with an authentic sample.³ Fraction **F-2** was also purified using an ODS column. Elution with MeOH–H₂O (4:1) of the column furnished abieta-8,11,13-trien-18-yl succinate, 20 mg, $[\alpha]^{23}_D+32^{\circ}$ (*c* 1.0, CHCl₃).²

7α,15-Dihydroxyabieta-8,11,13-trien-18-al (1): colorless oil; $[α]^{23}{}_D - 17^\circ$ (*c* 0.21, CHCl₃); IR (film) $ν_{max}$ 3380 (OH), 2971, 2931, 1717 (–CHO), 1498 and 1456 (aromatic ring) cm⁻¹; ¹H and ¹³C NMR, see Tables 1 and 2; EIMS (70 eV) *m/z* 316 [M]⁺ (12), 301 [M – Me]⁺ (100), 298 [M – H₂O]⁺ (36), 280 [M – 2H₂O]⁺ (9), 269 (16), 265 (15), 195 (20), 155 (14), 59 (12); HREIMS *m/z* 316.2044 (calcd for C₂₀H₂₈O₃, 316.2037).

Acetylation of Compound 1. A mixture of compound **1** (2 mg) in dried pyridine–Ac₂O (1:1, 1 mL) was left at room

temperature overnight. Workup as usual yielded a residue (3 mg), which was purified by preparative TLC (*n*-hexane–EtOAc, 3:1) to furnish a monoacetate (**1a**), 1.8 mg, as a colorless oil: $[\alpha]^{23}_{D}$ +21° (*c* 0.20, CHCl₃); IR (film) ν_{max} 3445 (OH), 2920, 2850, 1731 and 1238 (OAc), 1718 (–CHO), 1504 and 1463 (aromatic ring) cm⁻¹; ¹H and ¹³C NMR, see Tables 1 and 2; EIMS (70 eV) *m*/*z* 358 [M]⁺ (0.5), 340 (3), 316 (5), 298 (100), 286 (13), 255 (24), 173 (23), 155 (27), 141 (17), 59 (27), 43 (49); HREIMS *m*/*z* 358.2142 (calcd for C₂₂H₃₀O₄, 358.2142).

15,18-Dihydroxyabieta-8,11,13-trien-7-one (2): viscous oil; $[α]^{23}_D - 11^\circ$ (*c* 1.38, CHCl₃); UV (EtOH) $λ_{max}$ (log ε) 253 (3.96) and 299 (3.26) nm; IR (film) $ν_{max}$ 3408 (OH), 2972, 2932, 1668 (aryl C=O), 1497 and 1457 (aromatic ring), 1149, 983, 858 cm⁻¹; ¹H and ¹³C NMR, see Tables 1 and 2; EIMS (70 eV) m/z 316 [M]⁺ (6), 301 [M - Me]⁺ (100), 283 [M - Me - H₂O]⁺ (7), 203 (19), 187 (6), 115 (4), 43 (15); HREIMS m/z 316.2044 (calcd for C₂₀H₂₈O₃, 316.2037).

Acetylation of Compound 2. A mixture of compound 2 (20 mg) and dried pyridine-Ac₂O (1:1, 2 mL) was left at room temperature overnight. The reaction mixture was evaporated under reduced pressure to give a residue (23 mg), which showed two spots on TLC (*n*-hexane-EtOAc, 3:1). Si gel column chromatography of the residue yielded a monoacetate (2a), 12 mg, as a viscous oil: $[\alpha]^{23}_{D} - 12^{\circ}$ (*c* 1.06, CHCl₃); IR (film) v_{max} 3463 (OH), 2971, 2934, 1739 and 1238 (OAc), 1681 (aryl C=O), 1607, 1491 and 1459 (aromatic ring) cm⁻¹; ¹H and ¹³C NMR, see Tables 1 and 2; EIMS (70 eV) *m*/*z* 358 [M]⁺ (10), $343 [M - Me]^+$ (100), 298 (15), 283 (45), 265 (24), 203 (24), 187 (78); HREIMS *m*/*z* 358.2150 (calcd for C₂₂H₃₀O₄, 358.2142), and a diacetate (**2b**), 6 mg, as a viscous oil: $[\alpha]^{23}_{D} - 40^{\circ}$ (*c* 0.46, CHCl₃); IR (film) v_{max} 2935, 1737 and 1241 (OAc), 1683 (aryl C=O), 1610, 1492 and 1466 (aromatic ring) cm⁻¹; ¹H and ¹³C NMR, see Tables 1 and 2; EIMS (70 eV) *m*/*z* 400 [M]⁺ (10), 357 (41), 341 (97), 325 (39), 280 (44), 265 (72), 245 (24), 185 (29), 43 (100); HREIMS m/z 400.2250 (calcd for C₂₄H₃₂O₅, 400.2248).

18-*nor*-**4**,**15**-**Dihydroxyabieta**-**8**,**11**,**13**-**trien**-**7**-**one** (3): viscous oil; $[\alpha]^{23}_{D}$ +6° (*c* 1.2, CHCl₃); UV (EtOH) λ_{max} (log ϵ) 253 (3.94) and 297 (3.31) nm; IR (film) ν_{max} 3417 (OH), 2973, 2934, 1673 (aryl C=O), 1607, 1490 and 1457 (aromatic ring) cm⁻¹; ¹H and ¹³C NMR, see Tables 1 and 2; EIMS (70 eV) *m/z* 302 [M]⁺ (5), 287 [M - Me]⁺ (100), 269 [M - Me - H₂O]⁺ (6), 241 (5), 199 (12), 171 (5), 43 (14); HREIMS *m/z* 302.1889 (calcd for C₁₉H₂₆O₃, 302.1881).

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